

Package ‘STRINGdb’

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Type Package

Title STRINGdb (Search Tool for the Retrieval of Interacting proteins database)

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Description The STRINGdb package provides a user-friendly interface to the STRING protein-protein interactions database (<http://www.string-db.org>).

License GPL-2

Depends R (>= 2.14.0), png, sqldf, plyr, igraph, RCurl, plotrix, methods, RColorBrewer, gplots, hash

Suggests RUnit, BiocGenerics

biocViews Network

PackageStatus Deprecated

NeedsCompilation no

R topics documented:

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STRINGdb-package

STRINGdb (an R interface to <http://string-db.org>)

Description

The R package STRINGdb provides a convenient interface to the STRING protein-protein interactions database for the R/bioconductor users. Please look at the manual/vignette to get additional information and examples on how to use the package. STRING is a database of known and predicted protein-protein interactions. It contains information from numerous sources, including experimental repositories, computational prediction methods and public text collections. Each interaction is associated with a combined confidence score that integrates the various evidences. STRING is regularly updated, the latest version 9.05 contains information on 5 millions proteins from more than 1100 species. The STRING web interface is freely accessible at: <http://string-db.org/>

Details

Package: STRINGdb
Type: Package
Version: 1.0
Date: 2013-05-29
License: What license is it under?
Depends: methods

Author(s)

Andrea Franceschini

References

Franceschini, A (2013). STRING v9.1: protein-protein interaction networks, with increased coverage and integration. In: 'Nucleic Acids Res. 2013 Jan;41(Database issue):D808-15. doi: 10.1093/nar/gks1094. Epub 2012 Nov 29'.

See Also

<http://stitch-db.org>

Examples

```
library(STRINGdb)
data(diff_exp_example1)

# create a new STRING_db object
```

```

string_db <- STRINGdb$new()

# map to STRING
example1_mapped = string_db$map( diff_exp_example1, "gene", removeUnmappedRows = TRUE )

# get the best 200 hits
hits = example1_mapped$STRING_id[1:200]

# plot the STRING network png
string_db$plot_network( hits )

# plot a protein-protein enrichment graph of the best 1000 hits in order to see how the ppi signal is distributed
string_db$plot_ppi_enrichment( example1_mapped$STRING_id[1:1000] )

##### use the "halo" mechanism in STRING to distinguish between down and up-regulated genes #####

# filter by p-value and add a color column (i.e. green down-regulated genes and red for up-regulated genes)
example1_mapped_pval05 = string_db$add_diff_exp_color( subset(example1_mapped, pvalue<0.05), logFcColStr="logFC" )

# post payload information to the STRING server
payload_id = string_db$post_payload( example1_mapped_pval05$STRING_id, colors=example1_mapped_pval05$color )

# display a STRING network png with the "halo"
string_db$plot_network( hits, payload_id=payload_id )

##### compute enrichment in GO annotations #####
enrichmentGO = string_db$get_enrichment( hits, category = "Process", methodMT = "fdr", iea = TRUE )
enrichmentKEGG = string_db$get_enrichment( hits, category = "KEGG", methodMT = "fdr", iea = TRUE )
head(enrichmentGO, n=7)
head(enrichmentKEGG, n=7)

##### Use STRING in order to get homologous proteins in other species #####

# get the reciprocal best hits of the following protein in all the STRING species
string_db$get_homologs_bests(c("9606.ENSP00000365757"), symbets = TRUE)

# get the homologs of the following two proteins in the mouse (i.e. species_id=10090)
string_db$get_homologs(c("9606.ENSP00000365757", "9606.ENSP00000352336"), target_species_id=10090, bitscore_thr=0.5)

##### Retrieve protein-protein interactions #####

# get the neighbors (in the ppi graph) of a given protein(s)
string_db$get_neighbors(c("9606.ENSP00000365757", "9606.ENSP00000352336"))

# get the interactions in between the input proteins
string_db$get_interactions(c("9606.ENSP00000365757", "9606.ENSP00000352336"))

```

```
##### Find clusters of interactions #####  
  
# get clusters  
clustersList = string_db$get_clusters(example1_mapped$STRING_id[1:600])  
  
# plot first 4 clusters  
par(mfrow=c(2,2))  
for(i in seq(1:4)){  
  string_db$plot_network(clustersList[[i]])  
}
```

add_diff_exp_color *add_diff_exp_color*

Description

Take in input a dataframe containing a logFC column that reports the logarithm of the difference in expression level. Add a "color" column to the data frame such that strongly downregulated genes are colored in green and strong upregulated genes are in red. When the down or up-regulation is instead weak the intensity of the color gets weaker as well, accordingly.

Usage

```
## S4 method for signature 'STRINGdb'  
add_diff_exp_color(screen, logFcColStr="logFC" )
```

Arguments

| | |
|-------------|---|
| screen | Dataframe containing the results of the experiment (e.g. the analyzed results of a microarray or RNAseq experiment) |
| logFcColStr | name of the column that contains the logFC of the expression |

Value

vector containing the colors

Author(s)

Andrea Franceschini

```
add_proteins_description  
    add_proteins_description
```

Description

Add description coluns to the proteins that are present in the data frame given in input. The data frame must contain a column named "STRING_id".

Usage

```
## S4 method for signature 'STRINGdb'  
add_proteins_description(screen)
```

Arguments

| | |
|--------|---|
| screen | Dataframe containing the results of the experiment (e.g. the analyzed results of a microarray or RNAseq experiment) |
|--------|---|

Value

returns the same dataframe given in input with an additional columns containing a description of the proteins.

Author(s)

Andrea Franceschini

```
benchmark_ppi    benchmark_ppi
```

Description

benchmark a list of protein-protein interactions using pathways (e.g. KEGG). The function outputs a table where the interactions are mapped to KEGG and the number of TPs and FPs are counted.

Usage

```
## S4 method for signature 'STRINGdb'  
benchmark_ppi(interactions_dataframe, pathwayType = "KEGG", max_homology_bitscore = 60, precision_wir
```

Arguments

| | |
|------------------------|--|
| interactions_dataframe | a data frame containing the sorted interactions to be benchmarked. The data frame should have the following column names: proteinA, proteinB, score |
| pathwayType | category to use to benchmark the interactions (default KEGG) |
| max_homology_bitscore | filter out pairs of homologous proteins, having a similarity bitscore higher than this parameter |
| precision_window | define the size of the precision window (i.e. the window used to scan the sorted interactions data frame in order to compute the benchmark precision). At the beginning and at the end of the sorted interactions data frame, the window is automatically enlarged (at the beginning) and reduced (at the end) |
| exclude_pathways | Exclude the terms that should not be used for benchmarking. If this parameter is set to "blacklist", a black list to be used maintained by our group is automatically downloaded from our servers (otherwise it is possible to specify a vector with the terms that have to be excluded). |

Value

interactions data frame where the interactions are mapped to KEGG and the number of TPs and FPs are counted.

Author(s)

Andrea Franceschini

benchmark_ppi_pathway_view
benchmark_ppi_pathway_view

Description

Takes in input the results of the benchmark_ppi function, and constructs a new table that provides a view at the pathway level (i.e. it lists all the pathways to which the interactions belong)

Usage

```
## S4 method for signature 'STRINGdb'  
benchmark_ppi_pathway_view(benchmark_ppi_data_frame, precision_threshold=0.2, pathwayType = "KEGG")
```

Arguments

benchmark_ppi_data_frame data frame that comes out from the benchmark_ppi function.

precision_threshold threshold that specify where to stop taking considering the interactions in the sorted input data frame (the list is scanned until the precision goes below this value).

pathwayType the pathway category to use (KEGG by default)

Value

data frame containing the pathways (i.e. terms) of the input proteins. Several parameters are reported: pathway coverage (i.e. number of interactions in the list belonging to the pathway / maximum number of interactions between the pathay proteins (i.e. proteins * (proteins-1)/2)) total_representation (i.e. number of interactions in the list belonging to the pathway / size of the input data frame)

Author(s)

Andrea Franceschini

coeffOfvar

coeffOfvar

Description

coefficient of variation

Usage

coeffOfvar(x)

Arguments

x input number

Details

coefficient of variation

Value

coefficient of variation

Author(s)

Andrea Franceschini

| | |
|----------|-----------------|
| delColDf | <i>delColDf</i> |
|----------|-----------------|

Description

delete a column in the data frame

Usage

```
delColDf(df, colName)
```

Arguments

| | |
|---------|----------------------------------|
| df | data frame |
| colName | name of the column to be deleted |

Value

data frame

Author(s)

Andrea Franceschini

| | |
|-------------------|---|
| diff_exp_example1 | <i>example of microarray data (data processed from GEO GSE9008)</i> |
|-------------------|---|

Description

example of microarray data (data processed from GEO GSE9008)

Usage

```
data(diff_exp_example1)
```

Format

Data frames with 20861 observations on the following 3 variables.

| | |
|--------|--------------------|
| gene | a character vector |
| pvalue | a numeric vector |
| logFC | a numeric vector |

Source

Whyte L, Huang YY, Torres K, Mehta RG. Molecular mechanisms of resveratrol action in lung cancer cells using dual protein and microarray analyses. *Cancer Res* 2007.

downloadAbsentFile *downloadAbsentFile*

Description

download a file only if it is not present.

Usage

```
downloadAbsentFile(urlStr, oD = tempdir())
```

Arguments

| | |
|--------|-------------------------------------|
| urlStr | url from which to download the file |
| oD | directory where to store the file |

Author(s)

Andrea Franceschini

downloadAbsentFileSTRING
downloadAbsentFileSTRING

Description

download a STRING file only if it is not present or if it is corrupted.

Usage

```
downloadAbsentFileSTRING(urlStr, oD = tempdir())
```

Arguments

| | |
|--------|-------------------------------------|
| urlStr | url from which to download the file |
| oD | directory where to store the file |

Author(s)

Andrea Franceschini

| | |
|-------------|--------------------|
| get_aliases | <i>get_aliases</i> |
|-------------|--------------------|

Description

Loads and returns the STRING alias table.

Usage

```
## S4 method for signature 'STRINGdb'  
get_aliases( )
```

Value

a data frame containing the STRING alias table

Author(s)

Andrea Franceschini

| | |
|-----------------|------------------------|
| get_annotations | <i>get_annotations</i> |
|-----------------|------------------------|

Description

Loads and returns STRING annotations (i.e. GO annotations, KEGG pathways, domain databases). The annotations are stored in the "annotations" variable.

Usage

```
## S4 method for signature 'STRINGdb'  
get_annotations( )
```

Value

a data frame containing the annotations to the STRING proteins (e.g. GeneOntology, KEGG pathways, InterPro domains)

Author(s)

Andrea Franceschini

`get_annotatons_desc` *get_annotatons_desc*

Description

Returns a data frame with the description of every STRING annotation term (it downloads and caches the information the first time that is called).

Usage

```
## S4 method for signature 'STRINGdb'  
get_annotatons_desc()
```

Value

data frame with the description of every STRING annotation term.

Author(s)

Andrea Franceschini

`get_bioc_graph` *get_bioc_graph*

Description

Returns the interaction graph as an object of the graph package in Bioconductor.

Usage

```
## S4 method for signature 'STRINGdb'  
get_bioc_graph()
```

Value

interaction graph as an object of the graph package in Bioconductor.

Author(s)

Andrea Franceschini

get_clusters *get_clusters*

Description

Returns a list of clusters of interacting proteins. See the iGraph (<http://igraph.sourceforge.net/>) documentation for additional information on the algorithms.

Usage

```
## S4 method for signature 'STRINGdb'  
get_clusters(string_ids, algorithm="fastgreedy")
```

Arguments

`string_ids` a vector of STRING identifiers.
`algorithm` algorithm to use for the clustering. You can choose between "fastgreedy", "walk-trap", "spinglass" and "edge.betweenness").

Value

list of clusters of interacting proteins.

Author(s)

Andrea Franceschini

get_enrichment *get_enrichment*

Description

Returns the enrichment in pathways of the vector of STRING proteins that is given in input.

Usage

```
## S4 method for signature 'STRINGdb'  
get_enrichment(string_ids, category = "Process", methodMT = "fdr", iea = TRUE, minScore=NULL)
```

Arguments

| | |
|-------------------------|--|
| <code>string_ids</code> | a vector of STRING identifiers. |
| <code>category</code> | category for which to compute the enrichment (i.e. "Process", "Component", "Function", "KEGG", "Pfam", "InterPro"). The default category is "Process". |
| <code>methodMT</code> | method to be used for the multiple testing correction. (i.e. "fdr", "bonferroni"). The default is "fdr". |
| <code>iea</code> | specify whether you also want to use electronic inference annotations |
| <code>minScore</code> | with Tissue and Disease categories is possible to filter the annotations having an annotation score higher than this threshold (from 0 to 5) |

Value

Data frame containing the enrichment in pathways of the vector of STRING proteins that is given in input.

Author(s)

Andrea Franceschini

`get_graph`

get_graph

Description

Return an igraph object with the STRING network (for information about iGraph visit <http://igraph.sourceforge.net>)

Usage

```
## S4 method for signature 'STRINGdb'
get_graph()
```

Value

igraph object with the STRING network

Author(s)

Andrea Franceschini

References

Csardi G, Nepusz T: The igraph software package for complex network research, InterJournal, Complex Systems 1695. 2006. <http://igraph.sf.net>

See Also

In order to simplify the most common tasks, we do also provide convenient functions that wrap some iGraph functions. `get_interactions(string_ids)` # returns the interactions in between the input proteins `get_neighbors(string_ids)` # Get the neighborhoods of a protein (or of a vector of proteins) that is given in input. `get_subnetwork(string_ids)` # returns a subgraph from the given input proteins

| | |
|--------------|---------------------|
| get_homologs | <i>get_homologs</i> |
|--------------|---------------------|

Description

Returns the homologs of the given input identifiers that are present in the given `target_species_id`.

Usage

```
## S4 method for signature 'STRINGdb'  
get_homologs(string_ids, target_species_id, bitscore_threshold=NULL)
```

Arguments

`string_ids` a vector of **STRING** identifiers.
`target_species_id` NCBI taxonomy identifier of the species to query for homologs (the species must be present in the **STRING** database)
`bitscore_threshold` threshold on the bitscore of the blast alignment.

Value

Data frame containing the homologs of the given input identifiers and that are present in the given `target_species_id`.

Author(s)

Andrea Franceschini

`get_homologs_besthits` *get_homologs_besthits*

Description

Returns the best blast hits x species of the given input identifiers.

Usage

```
## S4 method for signature 'STRINGdb'
get_homologs_besthits(string_ids, symbets = FALSE, target_species_id = NULL, bitscore_threshold=NULL)
```

Arguments

`string_ids` a vector of STRING identifiers.
`target_species_id` NCBI taxonomy identifier of the species to query for homologs (the species must be present in the STRING database)
`bitscore_threshold` threshold on the bitscore of the blast alignment.
`symbets` specify whether you want only symmetrical best hits

Value

Data frame containing the best blast hits x species of the given input identifiers.

Author(s)

Andrea Franceschini

`get_interactions` *get_interactions*

Description

Shows the interactions in between the proteins that are given in input.

Usage

```
## S4 method for signature 'STRINGdb'
get_interactions(string_ids)
```

Arguments

`string_ids` a vector of STRING identifiers

Value

Data frame containing the interactions in between the input proteins.

Author(s)

Andrea Franceschini

| | |
|-----------------|-----------------|
| <i>get_link</i> | <i>get_link</i> |
|-----------------|-----------------|

Description

Returns a short link to the network page of our STRING website that shows the protein interactions between the given identifiers.

Usage

```
## S4 method for signature 'STRINGdb'  
get_link(string_ids, required_score=NULL, network_flavor="evidence", payload_id = NULL)
```

Arguments

| | |
|-----------------------------|--|
| <code>string_ids</code> | a vector of STRING identifiers. |
| <code>required_score</code> | minimum STRING combined score of the interactions (if left NULL we get the combined score of the object, which is 400 by default). |
| <code>network_flavor</code> | specify the flavor of the network ("evidence", "confidence" or "actions". default "evidence"). |
| <code>payload_id</code> | an identifier of payload data on the STRING server (see method <code>post_payload</code> for additional informations) |

Value

short link to the network page of our STRING website that shows the protein interactions between the input identifiers.

Author(s)

Andrea Franceschini

| | |
|---------------|----------------------|
| get_neighbors | <i>get_neighbors</i> |
|---------------|----------------------|

Description

Get the neighborhoods of a protein (or of a vector of proteins) that is given in input.

Usage

```
## S4 method for signature 'STRINGdb'
get_neighbors(string_ids)
```

Arguments

string_ids a vector of STRING identifiers

Value

vector containing the neighborhoods of a protein (or of a vector of proteins) that is given in input.

Author(s)

Andrea Franceschini

| | |
|---------|----------------|
| get_png | <i>get_png</i> |
|---------|----------------|

Description

Returns a png image of a STRING protein network with the given identifiers.

Usage

```
## S4 method for signature 'STRINGdb'
get_png(string_ids, required_score=NULL, network_flavor="evidence", file=NULL, payload_id=NULL)
```

Arguments

string_ids a vector of STRING identifiers.

required_score minimum STRING combined score of the interactions (if left NULL we get the combined score of the object, which is 400 by default).

network_flavor specify the flavor of the network ("evidence", "confidence" or "actions". default "evidence").

file file where to save the image

payload_id identifier of the payload

Value

Returns a png image of a STRING protein network with the given identifiers.

Author(s)

Andrea Franceschini

get_ppi_enrichment *get_ppi_enrichment*

Description

Returns a pvalue representing the enrichment in interactions of the list of proteins (i.e. the probability to obtain such a number of interactions by chance).

Usage

```
## S4 method for signature 'STRINGdb'  
get_ppi_enrichment(string_ids)
```

Arguments

string_ids a vector of STRING identifiers

Value

Returns a pvalue representing the enrichment in interactions of the list of proteins (i.e. the probability to obtain such a number of interactions by chance).

Author(s)

Andrea Franceschini

get_ppi_enrichment_full
 get_ppi_enrichment_full

Description

Returns a vector showing the enrichment in protein interactions in various positions of the list of genes in input. In practice, a list of 3 vectors is returned: 1) *enrichment* (i.e. enrichment computed in the window from 1 to x) 2) *enrichmentWindow* (i.e. enrichment computed in a sliding window of size determined by the "edgeWindow" parameters and the sliding steps determined by the "sliceWindow" parameter) 3) *enrichmentWindowExtended* (i.e. like the *enrichmentWindow*, but it also includes an initial window of size "windowExtendedReferenceThreshold" with respect to which to compute the enrichment)

Usage

```
## S4 method for signature 'STRINGdb'
get_ppi_enrichment_full(string_ids, sliceWindow = 20, edgeWindow = 140, windowExtendedReferenceThres
```

Arguments

`string_ids` a vector of STRING identifiers

`sliceWindow` defines the interval in proteins after which to compute the enrichment, scanning the list (i.e. the resolution)

`edgeWindow` size of the window that we use to compute the enrichment (i.e. the window pvalue is computed using the proteins inside this "edgeWindow")

`windowExtendedReferenceThreshold` defines the size of a window at the beginning of the list. The enrichment will be computed always including the proteins in this window

`growingWindowLimit` threshold where to stop the computation of the enrichment

Value

Returns a vector showing the enrichment in protein interactions in various positions of the list of genes in input.

Author(s)

Andrea Franceschini

| | |
|---------------------------|---------------------|
| <code>get_proteins</code> | <i>get_proteins</i> |
|---------------------------|---------------------|

Description

Returns the STRING proteins data frame. (it downloads and caches the information the first time that is called).

Usage

```
## S4 method for signature 'STRINGdb'
get_proteins()
```

Value

STRING proteins data frame.

Author(s)

Andrea Franceschini

| | |
|------------|-------------------|
| get_pubmed | <i>get_pubmed</i> |
|------------|-------------------|

Description

Returns vector with the PUBMED IDs of the publications that contain the names of the proteins in the input vector.

Usage

```
## S4 method for signature 'STRINGdb'  
get_pubmed(string_ids)
```

Arguments

string_ids a vector of STRING identifiers

Value

vector with the PUBMED IDs of the publications that contain the names of the proteins in the input vector.

Author(s)

Andrea Franceschini

| | |
|------------------------|-------------------------------|
| get_pubmed_interaction | <i>get_pubmed_interaction</i> |
|------------------------|-------------------------------|

Description

Returns vector with the PUBMED IDs of the publications that contain the names of both the input proteins.

Usage

```
## S4 method for signature 'STRINGdb'  
get_pubmed_interaction(STRING_id_a, STRING_id_b )
```

Arguments

STRING_id_a STRING identifier
STRING_id_b STRING identifier

Value

vector with the PUBMED IDs of the publications that contain the names of both the input proteins.

Author(s)

Andrea Franceschini

`get_STRING_species` *get_STRING_species*

Description

Returns a data frame with the species (i.e. organisms) that are present in STRING.

Usage

```
get_STRING_species(version=NULL, species_name=NULL)
```

Arguments

`version` STRING version
`species_name` name of the species that you are searching

Value

data frame with the species (i.e. organisms) that are present in STRING.

Author(s)

Andrea Franceschini

`get_subnetwork` *get_subnetwork*

Description

Returns the subgraph generated by the given input proteins.

Usage

```
## S4 method for signature 'STRINGdb'  
get_subnetwork(string_ids )
```

Arguments

`string_ids` a vector of STRING identifiers

Value

Returns the subgraph (i.e. an iGraph object) generated by the given input proteins.

Author(s)

Andrea Franceschini

| | |
|-------------|--------------------|
| get_summary | <i>get_summary</i> |
|-------------|--------------------|

Description

Returns a summary of the STRING sub-network containing the identifiers provided in input.

Usage

```
## S4 method for signature 'STRINGdb'  
get_summary(string_ids)
```

Arguments

string_ids a vector of STRING identifiers

Value

Returns a summary (i.e. a text description) of the STRING sub-network containing the identifiers provided in input.

Author(s)

Andrea Franceschini

| | |
|-------------------|--------------------------|
| get_term_proteins | <i>get_term_proteins</i> |
|-------------------|--------------------------|

Description

Returns the proteins annotated to belong to a given term.

Usage

```
## S4 method for signature 'STRINGdb'  
get_term_proteins(term_ids, string_ids=NULL, enableIEA=TRUE)
```

Arguments

| | |
|------------|---|
| term_ids | vector of terms |
| string_ids | a vector of STRING identifiers. If the variable is set, the method returns only the proteins that are present in this vector. |
| enableIEA | whether to consider also Electronic Inferred Annotations |

Value

Returns the proteins annotated to belong to a given term.

Author(s)

Andrea Franceschini

interactions_example *example of a protein-protein interactions sorted data frame*

Description

example of a sorted list of protein-protein interactions, resulta our cooccurrence algorithm (SVD_Phy)

Usage

```
data(interactions_example)
```

Format

Data frames with 20861 observations on the following 3 variables.

proteinA a character vector

proteinB a character vector

score a numeric vector

load

load

Description

Downloads and returns the STRING network (the network is set also in the graph variable of the STRING_db object).

It makes use of the variables: "backgroundV" vector containing STRING identifiers to be used as background (i.e. the STRING network loaded will contain only the proteins that are present also in this vector) "score_threshold" STRING combined score threshold (the network loaded contains only interactions having a combined score greater than this threshold)

Usage

```
## S4 method for signature 'STRINGdb'  
load()
```

Value

STRING network (i.e. an iGraph object. For info look to <http://igraph.sourceforge.net>)

Author(s)

Andrea Franceschini

load_all

load_all

Description

Force download and loading of all the files (so that you can later store the object on the hard disk if you like). It makes use of the variables: "backgroundV" vector containing STRING identifiers to be used as background (i.e. the STRING network loaded will contain only the proteins that are present also in this vector) "score_threshold" STRING combined score threshold (the network loaded contains only interactions having a combined score greater than this threshold)

Usage

```
## S4 method for signature 'STRINGdb'  
load_all()
```

Author(s)

Andrea Franceschini

| | |
|-----|------------|
| map | <i>map</i> |
|-----|------------|

Description

Maps the gene identifiers of the input dataframe to STRING identifiers. It returns the input dataframe with the "STRING_id" additional column.

Usage

```
## S4 method for signature 'STRINGdb'
map(my_data_frame, my_data_frame_id_col_names, takeFirst=TRUE, removeUnmappedRows=FALSE, quiet=FALSE)
```

Arguments

| | |
|---|--|
| <code>my_data_frame</code> | data frame provided as input. |
| <code>my_data_frame_id_col_names</code> | vector containing the names of the columns of "my_data_frame" that have to be used for the mapping. |
| <code>takeFirst</code> | boolean indicating what to do in case of multiple STRING proteins that map to the same name. If TRUE, only the first of those is taken. Otherwise all of them are used. (default TRUE) |
| <code>removeUnmappedRows</code> | remove the rows that cannot be mapped to STRING (by default those lines are left and their STRING_id is set to NA). |
| <code>quiet</code> | Setting this variable to TRUE we can avoid printing the warning relative to the unmapped values. |

Value

Returns the dataframe that is given in input with the "STRING_id" additional column.

Author(s)

Andrea Franceschini

| | |
|----|-----------|
| mp | <i>mp</i> |
|----|-----------|

Description

Maps the gene identifiers of the input vector to STRING identifiers (using a take first approach). It returns a vector with the STRING identifiers of the mapped proteins.

Usage

```
## S4 method for signature 'STRINGdb'
mp(protein_aliases)
```

Arguments

protein_aliases
vector of protein aliases that we want to convert to STRING identifiers

Value

It returns a vector with the STRING identifiers of the mapped proteins.

Author(s)

Andrea Franceschini

| | |
|--------------|---------------------|
| multi_map_df | <i>multi_map_df</i> |
|--------------|---------------------|

Description

mapping function (it add the possibility to map using more than one column of the data frame)

Usage

```
multi_map_df(dfToMap, dfMap, strColsFrom, strColFromDfMap, strColToDfMap, caseSensitive=FALSE)
```

Arguments

| | |
|-----------------|---|
| dfToMap | input data frame (that contains the columns that need to be mapped) |
| dfMap | data frame containing the mapping data |
| strColsFrom | sorted vector containing the names of the columns to be used in the input data frame for the mapping (the order of the elements in the vector defines the priority for the mapping) |
| strColFromDfMap | name of the column in the mapping data frame to be used as source for the mapping |
| strColToDfMap | name of the column in the mapping data frame to be used as target for the mapping |
| caseSensitive | specify whether the mapping should be case sensitive |

Value

data frame with an additional column containing the result of the mapping

Author(s)

Andrea Franceschini

| | |
|--------------|---------------------|
| plot_network | <i>plot_network</i> |
|--------------|---------------------|

Description

Plots an image of the STRING network with the given proteins.

Usage

```
## S4 method for signature 'STRINGdb'
plot_network(string_ids, payload_id=NULL, required_score=NULL, add_link=TRUE, add_summary=TRUE)
```

Arguments

| | |
|----------------|--|
| string_ids | a vector of STRING identifiers |
| payload_id | an identifier of payload data on the STRING server (see method post_payload for additional informations) |
| required_score | a threshold on the score that overrides the default score_threshold, that we use only for the picture |
| add_link | parameter to specify whether you want to generate and add a short link to the relative page in STRING. As default this option is active but we suggest to deactivate it in case one is generating many images (e.g. in a loop). Deactivating this option avoids to generate and store a lot of short-urls on our server. |
| add_summary | parameter to specify whether you want to add a summary text to the picture. This summary includes a p-value and the number of proteins/interactions. |

Author(s)

Andrea Franceschini

| | |
|---------------------|----------------------------|
| plot_ppi_enrichment | <i>plot_ppi_enrichment</i> |
|---------------------|----------------------------|

Description

Plots a graph showing the enrichment in protein interactions in various positions of the list of genes in input.

Usage

```
## S4 method for signature 'STRINGdb'
plot_ppi_enrichment(string_ids, file=NULL, sliceWindow = 20, edgeWindow = 140,
                    windowExtendedReferenceThreshold = 260, minVal=0.000000001, title="")
```

Arguments

| | |
|----------------------------------|---|
| string_ids | a vector of STRING identifiers |
| file | file where to save the graph as an image |
| sliceWindow | defines the interval in proteins after which to compute the enrichment, scanning the list (i.e. the resolution) |
| edgeWindow | size of the window that we use to compute the enrichment (i.e. the window pvalue is computed using the proteins inside this "edgeWindow") |
| windowExtendedReferenceThreshold | defines the size of a window at the beginning of the list. The enrichment will be computed always including the proteins in this window |
| title | title of the graph. |
| minVal | minimum value that the pvalue can assume in the log-scale graph. If the p-value is lower, we convert the value to this minimum value |
| quiet | if set to TRUE the method runs in quiet mode (turning off any output message) |

Author(s)

Andrea Franceschini

plot_ppi_enrichment_graph
plot_ppi_enrichment_graph

Description

Plots a graph showing the enrichment in protein interactions in various positions of the list of genes in input.

Usage

```
plot_ppi_enrichment_graph(proteins, ppi_network, file, sliceWindow, edgeWindow, windowExtendedRefere
```

Arguments

| | |
|-------------|--|
| proteins | a vector of protein identifiers |
| ppi_network | an igraph object containing the protein-protein interactions' graph. |
| file | file where to save the graph as an image |

| | |
|----------------------------------|---|
| sliceWindow | defines the interval in proteins after which to compute the enrichment, scanning the list (i.e. the resolution) |
| edgeWindow | size of the window that we use to compute the enrichment (i.e. the window pvalue is computed using the proteins inside this "edgeWindow") |
| windowExtendedReferenceThreshold | defines the size of a window at the beginning of the list. The enrichment will be computed always including the proteins in this window |
| title | title of the graph. |
| minVal | minimum value that the pvalue can assume in the log-scale graph. If the p-value is lower, we convert the value to this minimum value |
| quiet | if set to TRUE the method runs in quiet mode (turning off any output message) |

Author(s)

Andrea Franceschini

References

Pradines JR, Farutin V, Rowley S, Dancik V. Analyzing protein lists with large networks: edge-count probabilities in random graphs with given expected degrees. *J. Comput. Biol.* 2005;12:113-128. Franceschini, A et al. (2013). STRING v9.1: protein-protein interaction networks, with increased coverage and integration. In: *Nucleic Acids Res.* 2013 Jan;41(Database issue)

| | |
|--------------|---------------------|
| post_payload | <i>post_payload</i> |
|--------------|---------------------|

Description

Posts the input to STRING and returns an identifier that you can use to access the payload when you enter in our website.

Usage

```
## S4 method for signature 'STRINGdb'
post_payload(stringIds, colors=NULL, comments=NULL, links=NULL, iframe_urls=NULL, logo_imgF=NULL, leg
```

Arguments

| | |
|-----------|--|
| stringIds | vector of STRING identifiers. |
| colors | vector containing the colors to use for a every STRING identifier (the order of the elements must match those in the string_ids vector) |
| comments | vector containing the comments to use for every STRING identifier (the order of the elements must match those in the string_ids vector) |
| links | vector containing the links to use for every STRING identifier (the order of the elements must match those in the string_ids vector) |

| | |
|-------------|--|
| iframe_urls | vector containing the urls of the iframes to use for every STRING identifier (the order of the elements must match those in the string_ids vector). |
| logo_imgF | path to a file containing the logo image to be display in the STRING website |
| legend_imgF | path to a file containing a legend image to be display in the STRING website |

Value

identifier of the payload.

Author(s)

Andrea Franceschini

ppie.compLambda *ppie.compLambda*

Description

compute the number of expected interactions between a set of proteins.

Usage

```
ppie.compLambda(degrees, edgeNum)
```

Arguments

| | |
|---------|---|
| degrees | vector containing the degrees of the nodes of a set of proteins |
| edgeNum | total number of edges of the entire graph |

Value

number of expected interactions

Author(s)

Andrea Franceschini

References

Pradines JR, Farutin V, Rowley S, Dancik V. Analyzing protein lists with large networks: edge-count probabilities in random graphs with given expected degrees. *J. Comput. Biol.* 2005;12:113-128. Franceschini, A et al. (2013). STRING v9.1: protein-protein interaction networks, with increased coverage and integration. In: *Nucleic Acids Res.* 2013 Jan;41(Database issue)

`ppie.compLambdaL1L2` *ppie.compLambdaL1L2*

Description

Compute lambda L1L2

Usage

```
ppie.compLambdaL1L2(degreesI, degreesJ, edgeNum)
```

Arguments

| | |
|-----------------------|---|
| <code>degreesI</code> | vector containing the degrees of the nodes present in nodeSet I |
| <code>degreesJ</code> | vector containing the degrees of the nodes present in nodeSet J |
| <code>edgeNum</code> | total number of edges of the entire graph |

Details

compute the number of expected interactions between two sets of nodes

Author(s)

Andrea Franceschini

`ppie.compPij` *ppie.compPij*

Description

Compute the pvalue of protein I to interact with protein J (look at the Pradines paper referenced below)

Usage

```
ppie.compPij(degI, degJ, edgeNum)
```

Arguments

| | |
|----------------------|---|
| <code>degI</code> | degree of protein I |
| <code>degJ</code> | degree of protein J |
| <code>edgeNum</code> | total number of edges of the entire graph |

Value

return the pvalue of protein I to interact with protein J (look at the Pradines paper referenced below)

Author(s)

Andrea Franceschini

References

Pradines JR, Farutin V, Rowley S, Dancik V. Analyzing protein lists with large networks: edge-count probabilities in random graphs with given expected degrees. *J. Comput. Biol.* 2005;12:113-128. Franceschini, A et al. (2013). STRING v9.1: protein-protein interaction networks, with increased coverage and integration. In: *Nucleic Acids Res.* 2013 Jan;41(Database issue)

`ppie.getNumEdgesBetween`
ppie.getNumEdgesBetween

Description

find the number of interactions between two sets of nodes

Usage

`ppie.getNumEdgesBetween(graph, nodesFrom, nodesTo)`

Arguments

| | |
|------------------------|---------------|
| <code>graph</code> | igraph object |
| <code>nodesFrom</code> | list of nodes |
| <code>nodesTo</code> | list of nodes |

Value

return the number of interactions between two sets of nodes

Author(s)

Andrea Franceschini

`ppi_enrichment` *ppi_enrichment*

Description

Computes the enrichment in protein-protein interactions.

Usage

```
ppi_enrichment(hitList, ppi_network)
```

Arguments

| | |
|--------------------------|---|
| <code>hitList</code> | sorted list of proteins (from the most significant to the least significant) |
| <code>ppi_network</code> | an igraph object containing the graph of the protein-protein interaction's network. |

Value

| | |
|-------------------------|--|
| <code>enrichment</code> | pvalue that describes the probability to get such a number of interactions by chance |
| <code>lambda</code> | expected number of interactions |

Author(s)

Andrea Franceschini

References

Pradines JR, Farutin V, Rowley S, Dancik V. Analyzing protein lists with large networks: edge-count probabilities in random graphs with given expected degrees. *J. Comput. Biol.* 2005;12:113-128. Franceschini, A et al. (2013). STRING v9.1: protein-protein interaction networks, with increased coverage and integration. In: *Nucleic Acids Res.* 2013 Jan;41(Database issue)

`ppi_enrichment_full` *ppi_enrichment_full*

Description

Compute the enrichment in protein-protein interactions of a sorted list of proteins. The computation is repeated at different positions in the list.

Usage

```
ppi_enrichment_full(hitList, ppi_network, sliceWindow, edgeWindow, windowExtendedReferenceThreshold,
```

Arguments

| | |
|----------------------------------|--|
| hitList | sorted list of proteins (from the most significant to the least significant) |
| ppi_network | an igraph object containing the protein-protein interactions' graph. |
| sliceWindow | defines the interval in proteins after which to compute the enrichment, scanning the list (i.e. the resolution) |
| edgeWindow | size of the window that we use to compute the enrichment (i.e. the window pvalue is computed using the proteins inside this "edgeWindow") |
| windowExtendedReferenceThreshold | when we compute the "windowExtended" pvalue we are computing the pvalue that considers the following interactions: 1) the interactions inside the edgeWindow (as we do with the edgeWindow pvalue) 2) the interactions that connects the proteins in the edgeWindow with the proteins in another window at the beginning of the list (i.e. the windowExtendedReference). windowExtendedReferenceThreshold defines the size of this windowExtendedReference window. In this way we can compute, in a reliable way, the enrichment of a sorted list of proteins, in various positions of the list. |
| growingWindowLimit | stop to compute the enrichment (from position 1 to position n) after growingWindowLimit proteins in the sorted list. (this limit speeds up the computation of the 2 other types of enrichment) |
| quiet | if set to TRUE the method runs in quiet mode (turning off any output message) |

Value

| | |
|--------------------------|--|
| enrichment | vector containing the enrichments in protein-protein interactions of the input list of genes (the length of the vector is $\text{length}(\text{hitList})/\text{sliceWindow}$). |
| enrichmentWindow | vector containing the enrichments in protein-protein interactions of the input list of genes (the length of the vector is $\text{length}(\text{hitList})/\text{sliceWindow}$). The enrichment is computed considering only the proteins inside the sliding window |
| enrichmentWindowExtended | vector containing the enrichments in protein-protein interactions of the input list of genes (the length of the vector is $\text{length}(\text{hitList})/\text{sliceWindow}$). Look at the description of the windowExtendedReferenceThreshold variable |

Author(s)

Andrea Franceschini

References

Pradines JR, Farutin V, Rowley S, Dancik V. Analyzing protein lists with large networks: edge-count probabilities in random graphs with given expected degrees. *J. Comput. Biol.* 2005;12:113-128. Franceschini, A et al. (2013). STRING v9.1: protein-protein interaction networks, with increased coverage and integration. In: *Nucleic Acids Res.* 2013 Jan;41(Database issue)

```
remove_homologous_interactions  
    remove_homologous_interactions
```

Description

With this method it is possible to remove the interactions that are composed by a pair of homologous/similar proteins, having a similarity bitscore between each other higher than a threshold.

Usage

```
## S4 method for signature 'STRINGdb'  
remove_homologous_interactions(interactions_dataframe, bitscore_threshold = 60)
```

Arguments

```
interactions_dataframe  
    a data frame containing the sorted interactions to be benchmarked. The data  
    frame should have the following column names: proteinA, proteinB, score  
bitscore_threshold  
    filter out pairs of homologous proteins, having a similarity bitscore higher than  
    this parameter
```

Value

interactions data frame where the homologous pairs have been removed, from the input interactions' data frame

Author(s)

Andrea Franceschini

```
renameColDf    renameColDf
```

Description

Rename a column of a data frame

Usage

```
renameColDf(df, colOldName, colNewName)
```

Arguments

| | |
|-------------------------|---------------------------|
| <code>df</code> | input data frame |
| <code>colOldName</code> | column name to be changed |
| <code>colNewName</code> | new column name |

Value

data frame with the column name changed

Author(s)

Andrea Franceschini

| | |
|-----------------------------|-----------------------|
| <code>set_background</code> | <i>set_background</i> |
|-----------------------------|-----------------------|

Description

With this method you can specify a vector of proteins to be used as background. The network is reloaded and only the proteins that are present in the background vector are inserted in the graph. Besides, the background is taken in consideration for all the enrichment statistics.

Usage

```
## S4 method for signature 'STRINGdb'  
set_background(background_vector )
```

Arguments

| | |
|--------------------------------|--------------------------------------|
| <code>background_vector</code> | vector of STRING protein identifiers |
|--------------------------------|--------------------------------------|

Author(s)

Andrea Franceschini

STRINGdb

STRINGdb (an R interface to <http://string-db.org>)

Description

The R package STRINGdb provides a convenient interface to the STRING protein-protein interactions database for the R/bioconductor users. Please look at the manual/vignette to get additional information and examples on how to use the package. STRING is a database of known and predicted protein-protein interactions. It contains information from numerous sources, including experimental repositories, computational prediction methods and public text collections. Each interaction is associated with a combined confidence score that integrates the various evidences. STRING is regularly updated, the latest version 9.05 contains information on 5 millions of proteins from more than 1100 species. The STRING web interface is freely accessible at: <http://string-db.org/>

Usage

```
STRINGdb(...)
```

Arguments

```
...          description
```

Source

```
http://string-db.org
```

References

Franceschini, A (2013). STRING v9.1: protein-protein interaction networks, with increased coverage and integration. In: 'Nucleic Acids Res. 2013 Jan;41(Database issue):D808-15. doi: 10.1093/nar/gks1094. Epub 2012 Nov 29'.

Examples

```
library(STRINGdb)
data(diff_exp_example1)

# create a new STRING_db object
string_db <- STRINGdb$new()

# map to STRING
example1_mapped = string_db$map( diff_exp_example1, "gene", removeUnmappedRows = TRUE )

# get the best 200 hits
hits = example1_mapped$STRING_id[1:200]

# plot the STRING network png
string_db$plot_network( hits )
```

```
# plot a protein-protein enrichment graph of the best 1000 hits in order to see how the ppi signal is distributed
string_db$plot_ppi_enrichment( example1_mapped$STRING_id[1:1000] )

##### use the "halo" mechanism in STRING to distinguish between down and up-regulated genes #####

# filter by p-value and add a color column (i.e. green down-regulated genes and red for up-regulated genes)
example1_mapped_pval05 = string_db$add_diff_exp_color( subset(example1_mapped, pvalue<0.05), logFcColStr="logFC" )

# post payload information to the STRING server
payload_id = string_db$post_payload( example1_mapped_pval05$STRING_id, colors=example1_mapped_pval05$color )

# display a STRING network png with the "halo"
string_db$plot_network( hits, payload_id=payload_id )

##### compute enrichment in GO annotations #####
enrichmentGO = string_db$get_enrichment( hits, category = "Process", methodMT = "fdr", iea = TRUE )
enrichmentKEGG = string_db$get_enrichment( hits, category = "KEGG", methodMT = "fdr", iea = TRUE )
head(enrichmentGO, n=7)
head(enrichmentKEGG, n=7)

##### Use STRING in order to get homologous proteins in other species #####

# get the reciprocal best hits of the following protein in all the STRING species
string_db$get_homologs_bests(c("9606.ENSP00000365757"), symbets = TRUE)

# get the homologs of the following two proteins in the mouse (i.e. species_id=10090)
string_db$get_homologs(c("9606.ENSP00000365757", "9606.ENSP00000352336"), target_species_id=10090, bitscore_thr=0.5)

##### Retrieve protein-protein interactions #####

# get the neighbors (in the ppi graph) of a given protein(s)
string_db$get_neighbors(c("9606.ENSP00000365757", "9606.ENSP00000352336"))

# get the interactions in between the input proteins
string_db$get_interactions(c("9606.ENSP00000365757", "9606.ENSP00000352336"))

##### Find clusters of interactions #####

# get clusters
clustersList = string_db$get_clusters(example1_mapped$STRING_id[1:600])

# plot first 4 clusters
par(mfrow=c(2,2))
for(i in seq(1:4)){
  string_db$plot_network(clustersList[[i]])
}
```

```
}

```

STRINGdb-class

Class "STRINGdb"

Description

The R package STRINGdb provides a convenient interface to the STRING protein-protein interactions database for the R/bioconductor users. Please look at the manual/vignette to get additional information and examples on how to use the package. STRING is a database of known and predicted protein-protein interactions. It contains information from numerous sources, including experimental repositories, computational prediction methods and public text collections. Each interaction is associated with a combined confidence score that integrates the various evidences. STRING is regularly updated, the latest version 9.05 contains information on 5 millions proteins from more than 1100 species. The STRING web interface is freely accessible at: <http://string-db.org/>

Extends

All reference classes extend and inherit methods from "[envRefClass](#)".

Fields

annotations: Object of class `data.frame` ~~
 annotations_description: Object of class `data.frame` ~~
 graph: Object of class `igraph` ~~
 proteins: Object of class `data.frame` ~~
 speciesList: Object of class `data.frame` ~~
 species: Object of class `numeric` ~~
 version: Object of class `character` ~~
 input_directory: Object of class `character` ~~
 backgroundV: Object of class `vector` ~~
 score_threshold: Object of class `numeric` ~~

Methods

set_background(background_vector): ~~
 post_payload(stringIds, colors, comments, links, iframe_urls, logo_imgF, legend_imgF):
 ~~
 plot_network(string_ids, payload_id, required_score): ~~
 plot_ppi_enrichment(string_ids, file, sliceWindow, edgeWindow, windowExtendedReferenceThreshold, mi
 ~~


```

map(my_data_frame, my_data_frame_id_col_names, takeFirst, removeUnmappedRows, quiet):
  ~~
load(): ~~
get_term_proteins(term_ids, string_ids, enableIEA): ~~
get_summary(string_ids): ~~
get_subnetwork(string_ids): ~~
get_ppi_enrichment_full(string_ids, sliceWindow, edgeWindow, windowExtendedReferenceThreshold, grow):
  ~~
get_ppi_enrichment(string_ids): ~~
get_proteins(): ~~
get_png(string_ids, required_score, network_flavor, file, payload_id): ~~
get_neighbors(string_ids): ~~
get_link(string_ids, required_score, network_flavor, payload_id): ~~
get_interactions(string_ids): ~~
get_homologs_besthits(string_ids, symbets, target_species_id, bitscore_threshold):
  ~~
get_homologs(string_ids, target_species_id, bitscore_threshold): ~~
get_graph(): ~~
get_enrichment(string_ids, category, methodMT, iea): ~~
get_clusters(string_ids, algorithm): ~~
get_annotations_desc(): ~~
get_annotations(): ~~
load_all(): ~~
initialize(...): ~~
add_proteins_description(screen): ~~
add_diff_exp_color(screen, logFcColStr): ~~
show(): ~~

```

Author(s)

Andrea Franceschini

References

Franceschini, A (2013). STRING v9.1: protein-protein interaction networks, with increased coverage and integration. In: 'Nucleic Acids Res. 2013 Jan;41(Database issue):D808-15. doi: 10.1093/nar/gks1094. Epub 2012 Nov 29'.

See Also

<http://stitch-db.org>

Examples

```
showClass("STRINGdb")
```

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